

TECHNICAL SHEET

**BA045**

**HAEMATOXYLIN (HARRIS)**

**Formula**

**Reagent:**

Hematoxylin	5.000 gm
Ammonium or potassium alum	100.000 gm
Mercuric oxide	2.500 gm
Alcohol 95%	50.000 ml
Distilled water	1000.000 ml

**Quality Control:**

**Appearance:** Red coloured solution clear without any particles.

**Directions:**

1. Flame the slide and place in xylene for 3-4 minutes.Repeat xylene treatment with agitation.
2. Dip in 100% absolute alcohol for 30-60 seconds. Next ,dip in 90%, 80% and in 70% absolute alcohol. Wash in tap water and rinse in distilled water.
3. Stain in Haematoxylin (S034) for 3-5 minutes. Wash in tap water.
4. Dip in 0.5% (v/v) hydrochloric acid.
5. Rinse in tap water for 30-60 seconds.
6. Dip in dilute ammonia water till section appears blue.
7. Wash in tap water and then rinse in 95% alcohol.
8. Agitate in eosin solution for 10-60 seconds. Drain stain solution.
9. Dip slide in 70% alcohol for 30-60 seconds.
10. Place in 95% alcohol for 30-60 seconds.
11. Place in absolute alcohol (2 changes, 30-60 seconds each).
12. Place the slide twice in xylene for 30-60 seconds.
13. Drain excess xylene and mount on DPX or Canada balsam with a coverslip.

The first 2 steps of the procedure are collectively referred to in all staining procedures as "deparaffinize." Steps 3-8 are referred to as "staining". The last 5 steps are referred to in all staining methods as "dehydrate, clear, and mount".

**Precautions :**

1. For Laboratory Use.
2. Follow proper, established laboratory procedures in handling and disposing of infectious materials.

**Use:** Recommended as nuclear stain for Immunohistochemical and cytochemical staining.

**Quality Control:**

**Microscopy:**

Nuclei: Blue  
Cytoplasm: Grey or pale blue

**Storage:** Below 30°C.

**Packing:** 100 ml

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**Disclaimer:**

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